

1 ***Introduction to Themed Series on Intestinal Stem Cells and the NIDDK Intestinal***
2 ***Stem Cell Consortium***

3
4 Timothy C. Wang^{1*}, Martin G. Martin², Calvin J. Kuo³, Ophir D. Klein⁴, and Joyce
5 Niland⁵
6

7
8 ¹Division of Digestive and Liver Diseases, Columbia University Medical Center, New
9 York, NY 10032, USA. Email: tcw21@columbia.edu
10

11 ²Department of Pediatrics, Division of Gastroenterology and Nutrition, Mattel
12 Children's Hospital and the David Geffen School of Medicine at UCLA, University of
13 California Los Angeles, Los Angeles, California, 90095 USA.
14

15 ³Department of Medicine, Division of Hematology, Stanford University School of
16 Medicine, Stanford, CA 94305, USA.
17

18 ⁴Department of Orofacial Sciences and Program in Craniofacial Biology, Department
19 of Pediatrics and Institute for Human Genetics, UCSF, San Francisco, CA 94143, USA.
20

21 ⁵Department of Diabetes and Cancer Discovery Science, City of Hope Comprehensive
22 Cancer Center, Duarte, CA 91010, USA.
23

- 24
25 • Corresponding Author:
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40

41 This short piece serves to introduce our themed series entitled, "The Engineered
42 Gut: Use of Stem Cells and Tissue Engineering to Study Physiology and Disease."
43 Given the remarkable progress in our understanding of intestinal stem cells (ISCs)
44 over the last decade, it seems timely to review the topic of ISC in some depth.
45 Indeed, it has been approximately ten years since the first report of Lgr5 as an ISC
46 marker, and the creation of the Intestinal Stem Cell Consortium by the National
47 Institute of Diabetes and Digestive and Kidney Diseases (NIDDK). The ISC field
48 and the Intestinal Stem Cell Consortium are moving into the next phase of discovery;
49 consequently it is reasonable to take stock of where we are and touch on the many
50 challenges that await us.

51 Theories of the origins of intestinal epithelial cells date back to the 1950's and
52 1960's, with the discovery of DNA and DNA labeling studies in rodents using ³H-
53 thymidine. Proliferation was found not to be evenly distributed throughout the
54 intestinal epithelium, but largely restricted to the lower three-fourths of the crypt,
55 with a general flow upwards. The initial concept was that if dedicated ISC exist,
56 they would reside at the base of the proliferative column, immediately above the
57 Paneth cells[3]. Detailed studies using autoradiography and electronic microscopy
58 by Cheng and Leblond enumerated the various progenitors, and these investigators
59 eventually came up with the "Unitarian Theory", suggesting that all four main
60 epithelial cell types (columnar, enteroendocrine, goblet and Paneth) are derived
61 from the same precursor[5]. Importantly, they identified undifferentiated (granule
62 free) potential progenitors located in the lower intestinal crypts (at cell positions +1
63 to +9) that they believed represented stem cells. Interestingly, some of these Crypt
64 Base Columnar (CBC) cells were located below Paneth cells at the +1 position.
65 Around the same time, Potten and Cairns were working on radiation studies in the
66 small intestine, studying the segregation of DNA, and found that radiation induced
67 apoptosis in a number of progenitors, but led to retention of the ³H-thymidine DNA
68 label for more than 30 days by a subset of cells[11]. They went on to show that
69 these label-retaining cells are rare cells that reside between positions +2 to +10,
70 with a peak at +4 - +5; thus the concept of the label-retaining cell or + 4 cell was
71 developed[12]. As a result, for decades the general thinking was that the stem cell
72 compartment of the intestinal epithelium was achieved by a hierarchical age
73 structure, with rare, slowly dividing radioresistant cells at the top of the hierarchy.

74
75 However, our notions were dramatically changed with the discovery by Clevers and
76 Barker in 2007 of the Lgr5 cells[2], abundant in the lower crypts, with more rapid
77 division, consistent with many of the features of the Crypt Base Columnar (CBC)
78 cells first described by Leblond. The Lgr5-CreERT knock-in allele was able to
79 lineage trace all intestinal epithelial lineages in a mosaic fashion over a fairly short
80 time course, consistent with an active stem cell. However, in 2008 Sangiorgi and
81 Capecchi reported the existence of a Bmi1+ population located at the +4 position in
82 the intestinal crypts, that appeared to divide slowly, self-renew, and give rise to all
83 of the differentiated lineages[13]. This reignited the debate regarding CBC versus
84 the +4 ISC, which was later resolved somewhat with the notion of a 2 stem cell
85 model[9].

86

87 At this critical launching point for the ISC field, the Intestinal Stem Cell Consortium
88 or ISCC came together. The notion of such a consortium was formulated by the
89 NIDDK during their workshop on Local Influences on Health and Repair of
90 Intestinal Epithelium (March 25-26, 2008). Formally established in 2009, from
91 the beginning the ISCC has functioned as a highly collaborative, interactive team-
92 science initiative, funded by the NIDDK and the National Institute of Allergy and
93 Infectious Diseases (NIAID). The primary initial focus of the ISCC was to advance the
94 understanding of intestinal epithelial stem cell biology during development,
95 homeostasis, regeneration and disease.

96

97 In the first phase of the ISCC ('Gen 1', 2009-2014), there were 8 intestinal stem cell
98 research centers (**Fig. 1A**), geographically well distributed, and coordinated by the
99 ISCC Coordinating Center at the City of Hope. A major goal of the consortium has
100 been to accelerate ISC research through collaborations and rapid exchange of
101 information, and to expand the tools available to the research community as a whole.
102 The ISCC initially focused on major questions in stem cell biology, including the role
103 of the CBC versus the +4 ISC, and the possible utility of other stem cell markers, with
104 a major focus on the mouse intestine. The group collaborated on an RNAseq project
105 for different cell subsets, established common FACS methods for sorting intestinal
106 epithelial cells, and developed standardized nomenclature for intestinal in vitro
107 cultures.

108

109 During this first Gen1 version of the ISCC, there were several other major advances
110 in the field. First, the technique of growing intestinal organoids from intestinal
111 crypts and even single Lgr5+ cells using 3D-cultures in Matrigel with ENR media
112 were reported by Sato et al[14]. At around the same time, Ootani et al described a
113 technique for long-term culture of both intestinal epithelial and mesenchymal
114 structures using air-liquid interface[10]. Shortly thereafter, the group led by Jim
115 Wells from Cincinnati reported the ability to grow human intestinal organoids
116 (HIOs) from endodermal tissue derived from pluripotent iPS or ES cells. They
117 developed a robust methodology to direct differentiation of human pluripotent stem
118 cells into intestinal tissue in vitro through sequential growth factors that mimic
119 embryonic intestinal development[15]. Further refinements resulted in structures
120 with both crypt-villus epithelium and mesenchyme, able to be engrafted in vivo[18].
121 Finally, the scientific basis for the observation that R-spondin was a critical factor in
122 the growth of intestinal organoids was clarified with the report by several groups
123 that Lgr5 was a receptor for R-spondin, and also associated with Wnt receptors[7].
124 Thus, while Wnt molecules possibly derived from Paneth cells were initially
125 considered to be a major niche factor, greater attention turned to R-spondins and
126 their role in regulating ISCs.

127

128 Studies by the ISCC and other labs during this first five-year period laid the
129 groundwork for the second phase of the ISCC, so-called 'Gen 2' (2014-2019) (**Fig.**
130 **1B**). A number of new centers were added to the ISCC, resulting in a greater focus on
131 human tissues and human intestinal organoids (HIOs), as well as on mitigation of

132 radiation injury to the intestine. With extensive research and validation of Lgr5 as
133 an active stem cell marker, the initial two-stem cell model evolved into the concept
134 of “plasticity” by reserve stem cells in the intestinal crypts, and multiple populations
135 including enteroendocrine cells were shown to lineage trace in the setting of Lgr5
136 cell loss or injury [21, 17, 16, 4]. The role of reserve populations was shown to be
137 particularly important and/or clinically relevant under conditions of radiation
138 injury or infection. Increased focus on human tissues resulted in widespread use of
139 the HIO technology by the group, with the remarkable observations that engineered
140 HIOs derived from pluripotent stem cells are able to develop a functional enteric
141 nervous system[19]. An additional major accomplishment by the group was the use
142 of novel gain-of-function and loss-of-function technologies to make the remarkable
143 insight that R-spondin and Wnt ligands are not identical in their activity as
144 previously assumed, but rather that R-spondin is in fact the major driver of stem cell
145 self-renewal[22]. As a next step, given the growing data that cell types beyond the
146 Paneth cell can contribute to ISC function and behavior, the group has begun
147 additional collaborative studies to define the role of stromal cells that constitute the
148 ISC niche, with a particular focus on mesenchymal cells expressing Foxl1[1],
149 Grem1[20] and PDGFR α [8]. The group also has shifted into studying the importance
150 of matrix contributions to stem cell behavior[6].

151
152 As the ISCC looks to 2019 and beyond, it is clear that the ultimate vision is *to develop*
153 *novel therapies targeting intestinal stem cells and their supportive niche to regenerate*
154 *and rebuild the human intestine*. Thus, the consortium is establishing new priorities,
155 and increasingly seeking new ways to translate discoveries in the stem cell field to
156 the bedside. Many of these new priorities are highlighted in this “Themed Series” on
157 intestinal stem cells. This series consists of 9 mini-reviews (**Table 1**) that cover
158 many active and emerging areas of research on ISCs, most of which involve ISCC
159 researchers and authors.

160
161 The work by the ISCC continues to be based on a solid foundation of intestinal stem
162 cell biology. In the first several themed reviews, our authors discuss a “Comparison
163 of Mouse and Fly ISCs”, highlighting the tremendous insights in ISC biology that
164 derive from research in a simpler invertebrate species, *Drosophila melanogaster*.
165 The second themed review discusses the diversity of ISCs, which includes more
166 active ISCs (e.g. Lgr5+ cells) as well as the more quiescent and/or reserve
167 population. Much of the past work on ISC function and lineage relationships comes
168 from genetic lineage tracing using inducible Cre-drivers; however investigators have
169 increasingly recognized the limitation of models employing tamoxifen-induced Cre.
170 Further, bulk RNAseq analysis often fails to take into account the complexity and
171 heterogeneity of intestinal tissue. Thus, in our third themed review, our authors
172 explore the utility and contribution of single cell approaches (i.e. single cell RNAseq)
173 to our understanding of the intestinal epithelium and lineage relationships. In our
174 fourth article, we move from a solitary focus on the epithelium to the surrounding
175 mesenchyme, and our authors discuss the concept of the stem cell niche as it applies
176 to the mammalian intestine.

177

178 In our fifth themed review, our authors discuss the role of epigenetics in the
179 regulation of intestinal stem cells. Insights into pluripotent stem cells and genetic
180 reprogramming have clarified the tremendous importance of epigenetic regulation
181 in maintaining the stem cell state, while also providing for the potential for plasticity
182 by intestinal crypt cells. In our sixth themed review, we describe the development of
183 HIOs from pluripotent stem cells by many of the original investigators in this field,
184 and their potential for use in translational research. In our seventh review,
185 investigators discuss how intestinal organoids can be used in vitro to study the
186 regenerative responses to diverse forms of intestinal damage, thus providing
187 insights into newer approaches for treating or mitigating such injury. In our eighth
188 themed review, our teams of biologists and bio-engineers comment on the latest
189 approaches and strategies for regenerative medicine, and how they could be applied
190 to the intestinal epithelium. Finally, in our last review, our authors discuss a
191 possible roadmap for bringing ISC technology to the bedside, and how
192 bioengineered organs or cells can potentially achieve FDA approval for the
193 treatment of human disease.

194
195
196
197
198
199
200
201
202
203
204
205
206
207
208
209
210
211
212

213 **Table 1:**

214 **Articles in the Themed Series on Intestinal Stem Cells**

215

Topic
Introduction to ISCs and the Consortium
Comparison of Mouse and Fly ISCs
Diversity of Stem Cells: Active and Quiescent Populations
Single-cell Approaches to Studying the Intestinal Epithelium and Lineage Relationships
Stem Cell Niche is Defined by the Signals that Maintain and Control the Activity of the Stem Cells
Epigenetic Regulation of ISCs
Use of Organoids to Study Regenerative Responses to Intestinal Damage
Development of HIOs
Bioengineering Strategies for Regenerative Medicine
Translational/Clinical: Working towards FDA Approval

216

217

218
219
220
221
222
223
224
225
226
227
228
229
230
231
232
233
234
235
236
237
238
239
240
241
242
243
244
245
246
247
248
249
250
251
252
253
254
255
256
257

REFERENCES:

1. **Aoki R, Shoshkes-Carmel M, Gao N, Shin S, May CL, Golson ML, Zahm AM, Ray M, Wisner CL, Wright CV, Kaestner KH**, *Foxl1-expressing mesenchymal cells constitute the intestinal stem cell niche*. *Cell Mol Gastroenterol Hepatol*, 2016. **2**(3): p. 175-188. DOI: <https://doi.org/10.1016/j.jcmgh.2015.12.004>
2. **Barker, N, van Es JH, Kuipers J, Kujala P, van den Born M, Cozijnsen M, Haegebarth A, Korving J, Begthel H, Peters PJ, Clevers H**, *Identification of stem cells in small intestine and colon by marker gene Lgr5*. *Nature*, 2007. **449**(7165): p. 1003-7. DOI: <https://doi.org/10.1038/nature06196>
3. **Bjerknes M and Cheng H**, *Gastrointestinal Stem Cells. II. Intestinal stem cells*. *Am J Physiol Gastrointest Liver Physiol*, 2005. **289**: p. G381 - G387. DOI: <https://doi.org/10.1152/ajpgi.00160.2005>
4. **Buczacki SJ, Zecchini HI, Nicholson AM, Russell R, Vermeulen L, Kemp R, Winton DJ**, *Intestinal label-retaining cells are secretory precursors expressing Lgr5*. *Nature*, 2013. **495**(7439): p. 65-9. DOI: <https://doi.org/10.1038/nature11965>
5. **Cheng H and Leblond CP**, *Origin, Differentiation and Renewal of the Four Main Epithelial Cell Types in the Mouse Small Intestine. V. Unitarian Theory of the Origin of the Four Epithelial Cell Types*. *Am. J. Anat.*, 1974. **141**: p. 537-562. DOI: <https://doi.org/10.1002/aja.1001410407>
6. **Cruz-Acuña R, Quiros M, Farkas AE, Dedhia PH, Huang S, Siuda D, Garcia-Hernandez V, Miller AJ, Spence JR, Nusrat A, Garcia AJ**, *Synthetic hydrogels for human intestinal organoid generation and colonic wound repair*. *Nat Cell Biol*, 2017. **19**(11): p. 1326-1335. DOI: <https://doi.org/10.1038/ncb3632>
7. **de Lau W, Barker N, Low TY, Koo BK, Li VS, Teunissen H, Kujala P, Haegebarth A, Peters PJ, van de Wetering M, Stange DE, van Es JE, Guardavaccaro D, Schasfoort RB, Mohri Y, Nishimori K, Mohammed S, Heck AJ, Clevers H**, *Lgr5 homologues associate with the Wnt receptors and mediate R-spondin signaling*. *Nature*, 2011. **476**(7360): p. 293-7. DOI: <https://doi.org/10.1038/nature10337>
8. **Greicius G, Kabiri Z, Sigmundsson K, Liang C, Bunte R, Singh MK, Virshup DM**, *PDGFRα+ pericryptal stromal cells are the critical source of Wnts and RSPO3 for murine intestinal stem cells in vivo*. *Proc Natl Acad Sci U S A.*, 2018. **115**(14): p. E3173-E3181. DOI: <https://doi.org/10.1073/pnas.1713510115>
9. **Li L and Clevers H**, *Coexistence of quiescent and active adult stem cells in mammals*. *Science*, 2010. **327**(5965): p. 542-5. DOI: <https://doi.org/10.1126/science.1180794>
10. **Ootani A, Li X, Sangiorgi E, Ho QT, Ueno H, Toda S, Sugihara H, Fujimoto K, Weissman IL, Capecchi MR, Kuo CJ**, *Sustained in vitro intestinal epithelial*

- 258 culture within a Wnt-dependent stem cell niche. *Nat Med*, 2009. **15**(6): p. 701-
259 6. DOI: <https://doi.org/10.1038/nm.1951>
- 260 11. **Potten CS, Hume WJ, Reid P, Cairns J**, *The Segregation of DNA in Epithelial*
261 *Stem Cells*. *Cell*, 1978. **15**: p. 899-906.
- 262 12. **Potten CS, Owen G, and Booth D**, *Intestinal stem cells protect their genome*
263 *by selective segregation of template DNA strands*. *J. Cell Science*, 2002. **115**: p.
264 2381-2388.
- 265 13. **Sangiorgi E and Capecchi MR**, *Bmi1 is expressed in vivo in intestinal stem*
266 *cells*. *Nature genetics*, 2008. **40**(7): p. 915-20. DOI:
267 <https://doi.org/10.1038/ng.165>
- 268 14. **Sato T, Vries RG, Snippert HJ, van de Wetering M, Barker N, Stange DE,**
269 **van Es JH, Abo A, Kujala P, Peters PJ, Clevers H**, *Single Lgr5 stem cells build*
270 *crypt-villus structures in vitro without a mesenchymal niche*. *Nature*, 2009.
271 **459**(7244): p. 262-5. DOI: <https://doi.org/10.1038/nature07935>
- 272 15. **Spence JR, Mayhew CN, Rankin SA, Kuhar MF, Vallance JE, Tolle K,**
273 **Hoskins EE, Kalinichenko VV, Wells SI, Zorn AM, Shroyer NF, Wells JM,**
274 *Directed differentiation of human pluripotent stem cells into intestinal tissues*
275 *in vitro*. *Nature*, 2011. **470**(7332): p. 105-9. DOI:
276 <https://doi.org/10.1038/nature09691>
- 277 16. **Tetteh PW, Basak O, Farin HF, Wiebrands K, Kretzschmar K, Begthel H,**
278 **van den Born M, Korving J, de Sauvage F, van Es JH, van Oudenaarden A,**
279 **Clevers H**, *Replacement of Lost Lgr5-Positive Stem Cells through Plasticity of*
280 *Their Enterocyte-Lineage Daughters*. *Cell Stem Cell*, 2016. **18**(2): p. 203-13.
281 DOI: <https://doi.org/10.1016/j.stem.2016.01.001>
- 282 17. **van Es JH, Sato T, van de Wetering M, Lyubimova A, Yee Nee AN,**
283 **Gregorieff A, Sasaki N, Zeinstra L, van den Born M, Korving J, Martens**
284 **ACM, Barker N, van Oudenaarden A, Clevers H**, *Dll1+ secretory progenitor*
285 *cells revert to stem cells upon crypt damage*. *Nat Cell Biol*, 2012. **14**(10): p.
286 1099-104. DOI: <https://doi.org/10.1038/ncb2581>
- 287 18. **Watson CL, Mahe MM, Munera J, Howell JC, Sundaram N, Poling HM,**
288 **Schweitzer JI, Vallance JE, Mayhew CN, Sun Y, Grabowski G, Finkbeiner**
289 **SR, Spence JR, Shroyer NF, Wells JM, Helmrath MA**, *An in vivo model of*
290 *human small intestine using pluripotent stem cells*. *Nat Med*, 2014. **20**(11): p.
291 1310-4. DOI: <https://doi.org/10.1038/nm.3737>
- 292 19. **Workman MJ, Mahe MM, Trisno S, Poling HM, Watson CL, Sundaram N,**
293 **Chang CF, Schiesser J, Aubert P, Stanley EG, Elefanty AG, Miyaoka Y,**
294 **Mandegar MA, Conklin BR, Neunlist M, Brugmann SA, Helmrath MA,**
295 **Wells JM**, *Engineered human pluripotent-stem-cell-derived intestinal tissues*
296 *with a functional enteric nervous system*. *Nat Med*, 2017. **23**(1): p. 49-59. DOI:
297 <https://doi.org/10.1038/nm.4233>
- 298 20. **Worthley DL, Churchhill M, Compton JT, Tailor Y, Rao M, Si Y, Levin D,**
299 **Schwartz MG, Uygur A, Hayakawa Y, Gross S, Renz BW, Setlik W,**

300 **Martinez AN, Chen X, Nizami S, Lee HG, Kang HP, Caldwell JM, Asfaha S,**
301 **Westphalen CB, Graham T, Jin G, Nagar K, Wang H, Kheirbek MA, Kolhe**
302 **A, Carpenter J, Glaire M, Nair A, Renders S, Manieri N, Muthupalani S,**
303 **Fox JG, Reichert M, Giraud AS, Schwabe RF, Pradere JP, Walton K,**
304 **Prakash A, Gumucio D, Rustgi AK, Stappenbeck TS, Friedman RA,**
305 **Gershon MD, Sims P, Grikscheit T, Lee FY, Karsenty G, Mukherjee S,**
306 **Wang TC, *Gremlin 1 identifies a skeletal stem cell with bone, cartilage, and***
307 ***reticular stromal potential.* Cell, 2015. 160(1-2): p. 269-84. DOI:**
308 **<https://doi.org/10.1016/j.cell.2014.11.042>**
309 21. **Yan KS, Gevaert O, Zheng GXY, Anchang B, Probert CS, Larkin KA, Davies**
310 **PS, Cheng ZF, Kaddis JS, Han A, Roelf K, Calderon RI, Cynn E, Hu X,**
311 **Mandleywala K, Wilhelmy J, Grimes SM, Corney DC, Boutet SC, Terry JM,**
312 **Belgrader P, Ziraldo SB, Mikkelsen TS, Wang F, von Furstenberg RJ,**
313 **Smith NR, Chandrakesan P, May R, Chrissy MAS, Jain R, Cartwright CA,**
314 **Niland JC, Hong YK, Carrington J, Breault DT, Epstein J, Houchen CW,**
315 **Lynch JP, Martin MG, Plevritis SK, Curtis C, Ji HP, Li L, Henning SJ, Wong**
316 **MH, Kuo CJ, *Intestinal Enteroendocrine Lineage Cells Possess Homeostatic and***
317 ***Injury-Inducible Stem Cell Activity.* Cell Stem Cell, 2017. 21(1): p. 78-90 e6.**
318 **DOI: <https://doi.org/10.1016/j.stem.2017.06.014>**
319 22. **Yan KS, Janda CY, Chang J, Zheng GXY, Larkin KA, Luca VC, Chia LA, Mah**
320 **AT, Han A, Terry JM, Ootani A, Roelf K, Lee M, Yuan J, Li X, Bolen CR,**
321 **Wilhelmy, Davies PS, Ueno H, von Furstenberg RJ, Belgrader P, Ziraldo**
322 **SB, Ordonez H, Henning SJ, Wong MH, Snyder MP, Weissman IL, Hsueh**
323 **AJ, Mikkelsen TS, Garcia KC, Kuo CJ, *Non-equivalence of Wnt and R-spondin***
324 ***ligands during lgr5+ intestinal stem-cell self-renewal.* Nature, 2017.**
325 **545(7653): p. 238-242. DOI: <https://doi.org/10.1038/nature22313>**
326

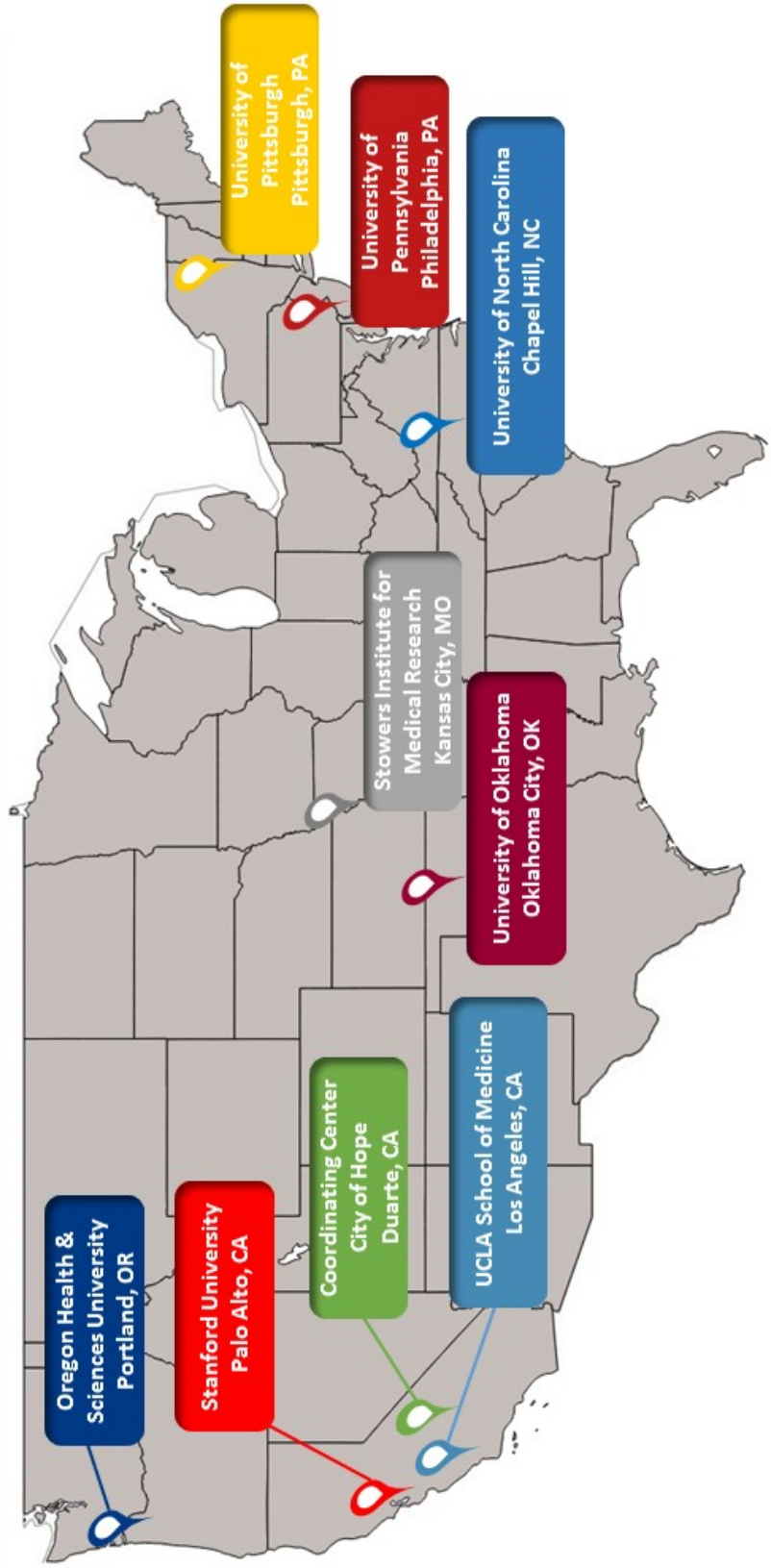
327 **Figure Legend**

328

329 Figure 1: Figures 1A and 1B: ISCC Institutions during “Gen 1” and “Gen 2

MAP of ISCC INSTITUTIONS 'GEN 1' 2009-2014

1A



MAP of ISCC INSTITUTIONS 'GEN 2' 2009-2014

1B

